Activation of Clustered IFNγ Target Genes Drives Cohesin-Controlled Transcriptional Memory

Graphical Abstract

Highlights
- IFNγ induces transcriptional memory for up to 14 days in cycling cells
- Transcription during priming increases probability to reinduce
- Changes in local chromatin structure at primed genes are maintained short term
- IFNγ-induced memory in gene clusters is locally controlled by cohesin

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In Brief
Siwek et al. find that IFNγ-induced transcriptional memory stably propagates through up to 14 cell division cycles. It manifests through an increased probability of primed cells to engage in target gene expression. The strongest memorized genes reside in two genomic clusters for which establishment of transcriptional memory is selectively restricted by cohesin.

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Article

Activation of Clustered IFNγ Target Genes Drives Cohesin-Controlled Transcriptional Memory

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SUMMARY

Cytokine activation of cells induces gene networks involved in inflammation and immunity. Transient gene activation can have a lasting effect even in the absence of ongoing transcription, known as long-term transcriptional memory. Here we explore the nature of the establishment and maintenance of interferon γ (IFNγ)-induced priming of human cells. We find that, although ongoing transcription and local chromatin signatures are short-lived, the IFNγ-primed state stably propagates through at least 14 cell division cycles. Single-cell analysis reveals that memory is manifested by an increased probability of primed cells to engage in target gene expression, correlating with the strength of initial gene activation. Further, we find that strongly memorized genes tend to reside in genomic clusters and that long-term memory of these genes is locally restricted by cohesin. We define the duration, stochastic nature, and molecular mechanisms of IFNγ-induced transcriptional memory, relevant to understanding enhanced innate immune signaling.

INTRODUCTION

Maintenance of gene expression states is essential for development and health. Local chromatin structure is implicated in preserving the transcriptional status of genes through cell division and cell lineages (Steffen and Ringrose, 2014). A role of cis-acting chromatin is most clearly defined in the maintenance of silent genes. During silencing, chromatin-associated proteins, modifications of DNA, and histones engage in self-propagating chromatin feedback loops that promote DNA compaction and restrict access for transcription factors (Gómez-Rodríguez and Jansen, 2013; Margueron and Reinberg, 2010). Active transcription can be maintained long term as well. However, in this case, self-propagation typically occurs through the self-amplified maintenance of trans-acting transcription factors that keep target genes expressed; for instance, during cellular differentiation (Grosschedl, 2013; Hoyler et al., 2012; Williams and Rudensky, 2007). Such cytoplasmic inheritance of transcription factors is arguably the most dominant form of epigenetic memory (Ptashne, 2011).

Whether cis-acting factors (e.g., local chromatin structure) contribute to maintenance of active transcriptional states remains unclear. Uncoupling the mechanisms that contribute to maintenance of transcription from those that contribute to transcription itself is challenging. However, there are instances of transcriptional memory where priming of gene activation can be maintained in the absence of the initial signal and, importantly, in the absence of ongoing transcription. Examples include sugar metabolism in Saccharomyces cerevisiae (Acar et al., 2005; Zacharioudakis et al., 2007), ecdysone response in Drosophila melanogaster (Pascual-García et al., 2017), heat response in Arabidopsis thaliana (Länkte et al., 2016), and nuclear transfer in Xenopus laevis (Ng and Gurdon, 2008). In all of these cases, a primed state of transcription is maintained after the initial signal subsides.

An emerging paradigm for long-term transcriptional memory in mammalian cells is the primed response to cytokines (D'Urso and Brickner, 2017), which results in transient but reversible expression of pro-inflammatory and innate immune genes (Kamada et al., 2018; Light et al., 2013). When primed, cells maintain a memory of interferon exposure even in the apparent absence of target gene expression. This poised state is revealed upon a second interferon pulse, resulting in enhanced expression of a subset of genes (Gialitakis et al., 2010; Light et al., 2013). Therefore, interferon signaling offers an opportunity to dissect the mechanisms underlying memory of transcription and identify local chromatin-based contributors to memory.

Moreover, interferon-induced transcriptional memory in mammals may relate to the broader physiological phenomenon of trained immunity. This is an adaptive form of innate immunity where an organism, when exposed to a pathogen and triggering an innate immune response, retains a poised physiological state for weeks or
months, resulting in an enhanced reaction upon a second exposure to the same or even entirely distinct insult (Netea et al., 2020). Striking examples of this phenomenon include enhanced resistance to Staphylococcus aureus after fungus-derived glucan treatment (Di Luzio and Williams, 1978; Marakalala et al., 2013) or hyperactivated anti-microbial effector genes after priming of macrophages with lipopolysaccharide (LPS) (Foster et al., 2007).

Interferon-mediated transcriptional memory has direct implications for enhanced innate immunity at the cell-autonomous level (e.g., resulting in an enhanced response to intracellular pathogens; Kamada et al., 2018; Sturge and Yarovinsky, 2014) and at the organismal level (Yao et al., 2018). Maintenance of a poised state to interferon may be one of the underlying mechanisms explaining trained immunity, and understanding the molecular nature of long-term transcriptional memory is therefore critical to advance our understanding of memory of innate immunity. However, studying transcriptional memory in the context of immunity poses challenges. For instance, priming of macrophages, key players in innate immunity, results not only in transient gene activation but also in sustained rewiring of transcriptional programs, enhancer activity, and lineage-specific transcription factor activation (Kang et al., 2017; Ostuni et al., 2013; Qiao et al., 2016). Therefore, in a physiological context, it is difficult to distinguish transient poised states from cellular differentiation. Interferon γ (IFNγ)-induced transcriptional memory has been established previously in HeLa cells. By using a non-hematopoietic cell type, we can avoid the confounding effects of lineage-specific transcription factor activation and therefore uncouple IFN-induced gene expression and memory from cellular differentiation.

Early work showed that a specific target gene, HLA-DRA, displays enhanced IFNγ-induced expression in cells that were primed previously by the same cytokine. Histone H3K4 dimethylation and retention of RNA polymerase II (RNA PolII) on promoters have been identified as molecular signatures that are retained in chromatin for up to 2 days after priming (Gialitakis et al., 2010; Light et al., 2013). Additionally, Nup98, a component of the nuclear pore (Light et al., 2013), as well as the CDK6+ mediator complex (D’Urso et al., 2016) are implicated in IFNγ transcriptional memory. More recently, IFNγ and IFNγ priming in mouse fibroblast and macrophages, respectively, has been shown to cause a similar memory effect. In this case, retention of the H3.3 replacement histone variant and histone H3K36 trimethylation were reported to be maintained on memory genes after removal of the cytokines (Kamada et al., 2018). However, in this context, retention of promoter-bound RNA polymerase was not detected.

Despite identification of several chromatin-associated factors required for transcriptional memory, there are many unanswered questions. Here we define the nature of IFNγ-primed genes and the duration of memory. We address the population dynamics of memory and the role of active transcription and local chromatin structure in maintenance of the primed state.

**RESULTS AND DISCUSSION**

**IFNγ Primes GBP5 for Long-Term Transcriptional Memory**

We aimed to identify all human genes that show long-term transcriptional memory following IFNγ priming in HeLa cells. Memory is defined as enhanced expression of target genes upon a second exposure to IFNγ (conceptualized in Figure 1A). We primed HeLa cells with IFNγ and allowed cells to recover for 48 h before reinduction to IFNγ (Figure 1B). In agreement with previous reports (Gialitakis et al., 2010; Light et al., 2013), we found HLA-DRA to be expressed 5-fold higher in IFNγ-primed relative to naive cells upon IFNγ re-exposure (Figure S1A). We used this IFNγ priming and reinduction regimen to unbiasedly discover all genes that behave in a manner analogous to HLA-DRA. We treated HeLa cells with IFNγ as in Figure 1B and analyzed transcript levels in naive cells, cells undergoing priming, primed, and reinduced cells by RNA sequencing. As expected, when comparing priming with naive cells, we observed numerous IFNγ-induced gene expression changes: activation (3,531 genes above 0.5 log2 fold change cutoff, adjusted p value [p-adj] < 0.05) and inhibition (2,117 genes below −0.5 log2 fold change cutoff, p-adj < 0.05) (STAR Methods; Figure S1B; Table S1). Importantly, we identified a small subset of IFNγ-induced genes that show transcriptional memory (as defined in Figure 1A) when comparing reinduced with priming transcriptomes (28 genes above 0.5 log2 fold change cutoff, p-adj < 0.05) (Figure 1C; Table S2), indicating that memory is not simply a consequence of signaling.

Among these are the previously established HLA-DRA gene (Figure S1A; Gialitakis et al., 2010; Light et al., 2013) and other major histocompatibility complex (MHC) class II genes. We found the gene encoding guanylate binding protein 5 (GBP5), an activator of inflammasome assembly (Shenoy et al., 2012), to display the strongest reinduction following initial priming (Figure 1C). We validated this observation by RT-qPCR measurements of GBP5 mRNA and determined an ~10-fold increase in expression of this target during reinduction relative to priming (Figure 1D). Enhanced GBP5 expression upon stimulation of primed cells was also detected at the protein level by western blot (Figure 1E). Importantly, long-term transcriptional memory is not restricted to cancer cells because we observed the same phenomenon in primary male fibroblasts (Figure 1E). However, priming is not universal among cultured human cells because HCT116 colon cancer cells and non-transformed retinal pigment epithelium (RPE-1) cells did not show transcriptional memory of GBP5 despite responding to IFNγ (Figure S1C). This suggest that, although IFNγ signaling and memory are broad phenomena, they are not hard wired, at least for GBP5, indicating that components involved in propagation of transcriptional memory are subject to regulation.

**Memory of Prior IFNγ: Induction Is Reversible but Persists for up to 14 Days in Continuously Cycling Cells**

One of the key features of transcriptional memory is its reversibility because this distinguishes it from a permanently altered transcriptional profile, such as cellular differentiation. Previous efforts have shown that priming by IFNγ lasts for at least 48 h (Gialitakis et al., 2010; Light et al., 2013) or even up to 8 days following priming by IFNγ in mouse cells (Kamada et al., 2018). However, the full extent of memory and the timing of its eventual loss have not been determined. To this end, we primed HeLa cells with IFNγ for 24 h and left cells to proliferate for an increasing number of days before reinduction (Figure 2A).
Enhanced GBP5 protein expression upon reinduction was evident for up to 14 days after priming but was lost at later time points (Figure 2B). During this time, cells are doubling daily and are passaged continuously, equaling up to $2^{14}$ cells. Similarly, GBP5 and HLA-DRA mRNA levels followed this trend; we detected enhanced activation for at least 7 days after priming (Figure 2C). This indicates that memory persists through many mitotic cell division cycles, at least for the GBP5 and HLA-DRA genes. Such mitotic stability of the primed state is unlikely to be a consequence of passive dilution of a factor induced during priming. Instead, an active positive feedback mechanism may be at play to maintain memory over such timescales. Importantly, the primed state is ultimately lost by 3 weeks of culture, indicating that priming is not a differentiation state involving rewiring of the transcriptional network.

more cells participating in transcription upon reinduction (hypothesis 2). We found no effect of IFNγ on selective cell proliferation or lethality, excluding a trivial explanation of cell selection (Figures S1D and S1E). To distinguish between the possibilities in Figure 3A, we performed single-cell RNA sequencing (RNA-seq) during the course of IFNγ induction and reinduction (Figure 3B). Consistent with the bulk RNA-seq data, single-cell mRNA levels showed that, on average, IFNγ priming resulted in enhanced expression of the GBP5 and HLA-DRA genes upon second exposure, whereas a constitutive control gene, ACTB, remained unaltered throughout the experiment (Figures 3C, S1F, and S1G). However, induction and reinduction were not uniform across the population. The per-cell transcriptional output of GBP5 was increased upon re-exposure to IFNγ, but only in 10 of 91 cells was expression boosted beyond the level
observed during priming (Figure 3C, labeled with red dots); in many cells, GBP5 was not detectably induced. To further illustrate this, we replotted the data in Figure 3C by binning cells based on their GBP5 expression level. In ~52% of cells, GBP5 was not induced during priming, whereas non-responsive cells represented only a minor (~15%) fraction following reinduction (Figure 3D). These data also indicate that stronger re-expression of GBP5 did occur to some extent. Approximately 35% of total transcriptional output comes from cells expressing higher levels during reinduction than during priming (red cells in Figure 3C), consistent with hypothesis 1. However, an increase in the population of cells that express GBP5 (hypothesis 2) appears to be responsible for the majority of the enhanced overall GBP5 output upon reinduction, constituting long-term transcriptional memory.

To explore this phenomenon further, we inserted an EGFP-based promoter trap at one of the genomic HeLa GBP5 alleles that allowed us to monitor GBP5 output on a per-cell basis (Figure 3E). We confirmed by flow cytometry that EGFP::GBP5 expression is specifically induced by IFNγ but not IFNα or IFNβ, as reported previously for GBP5 (Degrandi et al., 2007; Krapp et al., 2016). Priming of EGFP::GBP5 cells resulted in enhanced reinduction of EGFP by IFNγ, confirming preservation of transcriptional memory for the EGFP-tagged allele and the wild-type GBP5 allele in the same cells, as measured by western blot (Figure 3F). Cytometry-based analysis of cells showed that, during priming, 65% of cells detectably responded to IFNγ, whereas this number increased substantially to 80% upon reinduction (Figure 3G). Notably, the per-cell EGFP fluorescence did not increase significantly. These results are consistent with the single-cell RNA-seq data and indicate that, although transcriptional output per cell may be enhanced to some extent, the dominant mechanism of IFNγ priming of GBP5 is to render transcription upon second expose more likely.

Establishment of GBP5 Transcriptional Memory Correlates with Local Transcriptional Output

The population dynamics of GBP5 expression, as revealed by cytometry in Figure 3G, indicated that not all cells express, even upon repeated activation. We consider two possibilities. (1) cells have an inherent variability in their GBP5 response to IFNγ. For instance, this may be the case when there are clonal genetic differences in the cell population. (2) All cells have an equal but limited probability to induce GBP5, and exposure to IFNγ shifts this probability toward an increased chance of expressing upon later exposure. To explore these possibilities, we tracked cell populations that responded to priming or not and monitored their behavior during a second stimulation. Single-cell RNA-seq data of IFNγ-treated cells (Figure 3B) showed that all cells in the population (even when not activating EGFP::GBP5) were uniformly responsive to IFNγ (Figure S2A), indicating that the variability in GBP5 activation was not due to lack of IFNγ-mediated Janus kinase / Signal Transducer and
Figure 3. Priming Results in Increased Frequency of Activation and Enhanced GBP5 Expression upon Reinduction

(A) Alternative solutions for achieving IFNγ transcriptional memory.
(B) Scheme describing a single-cell RNA-seq transcriptional memory experiment to distinguish between the alternatives from (A).
(C) Representation of the single-cell RNA-seq data from HeLa cells for the GBP5 gene from the experiment shown in (B). Each dot represents the expression level of the GBP5 gene for one cell in the naive (N = 90), priming (N = 89), and reinduction state (N = 91).
(D) Binned representation of the single-cell RNA-seq data for the GBP5 gene from the experiment shown in (C).

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Ongoing Transcription Is Not Required for Maintenance of IFNγ Transcriptional Memory

One possible mechanism for the memory of transcription through successive cell division cycles is a continued low-level expression of the target genes, which is contrary to the pre-conception that inactivation is tightly shut down after IFNγ removal. To address this, we measured long-term transcriptional output of memory genes 2, 7, 14, 21, and 28 days following priming (Figure 4A). Although overall transcription levels are exceedingly low during the window of memory compared with the priming stage, we could still detect 2- and 3-fold higher levels of GBP5 and HLA-DRA mRNA, respectively, after 2 days compared with the low basal level of naive cells (Figure 4B). However, by day 7, we could not detect any difference, and transcript levels fully returned to baseline despite continued memory of GBP5 and HLA-DRA priming (Figure 2). We confirmed these observations in the EGFP::GBP5 reporter line, where we observed low but detectable expression of EGFP above the naive baseline 2 days but not 7 days following cytokine removal (Figure S3A).

Although we do not find evidence of continued target gene expression during memory, it is possible that the primed state of memory genes is maintained by continued expression of secondary genes that may drive reinduction upon IFNγ exposure. We re-analyzed the RNA-seq dataset as in Figure 1C but now plotted the differential expression of genes in primed versus naive cells (Figure 4C). In this scenario, most IFNγ-induced genes had returned to baseline. However, we did find a small subset of genes whose expression was still detectable 48 h after priming (93 genes above 0.5 log2 fold change cutoff, p-adj < 0.05) (Table S3), although the levels were dramatically lower than immediately following IFNγ induction (Figure S3B). Among these are HLA-DRA, as evident in Figure 4B (2 days after IFNγ); CD74, a chaperone stabilizing the peptide-free MHC class II complexes (Hiltbold and Roche, 2002); SERPING1, an inhibitor of the C1 complement pathway (Davis, 2004); and CXCL9, a chemokine involved in the inflammatory response (Egesten et al., 2007). However, none of these have an ascribed role in IFNγ signaling or target gene expression, making a continued transcription feedback loop driving memory unlikely.

To further test whether continued transcription is required to maintain memory, we inhibited transcription during the memory window, after priming but before reinduction (Figure 4D), using a brief treatment with triptolide, a fast-acting RNA polymerase II inhibitor (Vispé et al., 2009). We titrated the minimal amount needed to effectively block transcription to avoid non-specific effects (Figures S3 C and S3D) and treated cells transiently for 4 h during the primed state (Figure 4D). RT-qPCR analysis of GBP5 expression throughout the experiment showed that disruption of transcription had no bearing on GBP5 reinduction (Figure 4E). We conclude that continuous global transcriptional output is not required for maintenance of transcriptional memory of GBP5. These observations indicate that other mechanisms are likely responsible for maintenance of the primed state.

Short-Term Maintenance of H3.3, H3K4me2, and H3K79me2 on GBP Genes Following IFNγ Priming

We next determined whether chromatin features contribute to GBP5 transcriptional memory in the absence of ongoing transcription. We analyzed chromatin structure and protein occupancy in naive cells during priming and at different time points after IFNγ washout in primed cells to assess chromatin maintenance (Figure 5A). We employed a HeLa cell line expressing

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(E) Scheme outlining the GBP5 promoter trap cell line, in which EGFP was inserted into exon 2 downstream of the GBP5 translation start site. One allele was targeted, and the remaining allele remained unperturbed. Purple, 5' UTR; blue, GBP5 coding sequence.
(F) EGFP::GBP5 cells were subjected to the IFNγ treatment regimen outlined in Figure 1B, processed for fluorescence western blotting, and probed for GBP5 and EGFP expression. α-TUB, loading control. Tubulin-normalized fluorescence intensities are plotted.
(G) EGFP::GBP5 cells were subjected to the IFNγ treatment regimen as outlined in (B) and processed for cytometry. Cell frequencies as a function of EGFP fluorescence intensity are plotted. Red dotted lines are fiducial marks based on untagged cells and are used to define the cutoff for cell percentages expressing or not expressing.

See also Figure S2.
a SNAP-hemagglutinin (HA)-tagged version of the H3.3 variant to assess H3.3 levels (Bodor et al., 2013; Ray-Gallet et al., 2011). We confirmed transcriptional memory of the GBP5 gene 2 and 7 days after priming (Figure 5B), although the degree of long-term priming appears to be less pronounced compared with unmodified HeLa cells (Figures 1E and 2B). Using chromatin immunoprecipitation coupled to sequencing (ChIP-seq), we determined the occupancy of H3.3-HA (using the HA epitope) as well as RNA Pol II, histone H3 acetylation of lysine 27 (H3K27ac), trimethylation of lysine 36 (H3K36me3), dimethylation of lysine 79 (H3K79me2), and dimethylation of lysine 4 (H3K4me2) genome-wide. In addition, we determined global chromatin accessibility using ATAC-seq (Chen et al., 2016). As expected from active chromatin features, RNA Pol II, H3.3, and the above histone modifications accumulated during priming at GBP5 and the neighboring GBP4 gene (Figure 5C), correlating with opening of chromatin at the GBP5 and GBP4 promoters and induction of gene expression (Figures 1C–1E). However, we did not detect any appreciable maintenance of RNA Pol II, accessible chromatin, H3K27ac, or H3K36me3 on these genes (Figure 5C). Similarly, lack of RNA Pol II maintenance has been reported recently for IFNγ priming in mouse macrophages (Kamada et al., 2018). In contrast, we found low levels of H3.3-HA, H3K79me2, and H3K4me2 chromatin marks to be maintained on promoters of GBP genes for up to 2 days after IFNγ washout. Specifically, 2 days after priming, we detected low levels of H3.3-HA on the promoter of GBP5 but not GBP4 and H3K79me2 on the promoter of GBP5 but not GBP4, and H3K4me2 for both genes (Figures 5C and 5D). A role of H3.3 has been suggested previously for IFNγ transcriptional memory in mouse macrophages along with trimethylation of lysine 36 (Kamada et al., 2018), although we did not find significant retention of the latter mark in primed cells. H3K4me2 is of interest because it has been reported independently in several studies (Gialitakis et al., 2010; Light et al., 2013). Marking active promoters, it possibly plays a role in maintenance of a poised state, at least in the short term. In line with such a role, this modification has also been shown to be important in Dictyostelium discoideum gene expression memory (Muramoto et al., 2010) and mitotic inheritance of gene expression states in Xenopus laevis nuclear transfer experiments (Ng and Gurdon, 2008). Moreover, a mechanism of maintenance of this mark involving the SET3C methyltransferase complex has been demonstrated in heritable maintenance of a poised INO1 gene in yeast (D’Urso et al., 2016). Nevertheless,
the short maintenance of these chromatin features suggests that, although they may contribute to maintenance of the primed state, they are not the sole factors and unlikely to be key factors driving memory of IFN\(\gamma\) priming, at least for GBP genes.

**The Two Major Classes of Genes that Are Primed for Re-expression, GBPs and HLAs, Reside in Genomic Gene Clusters**

Analysis of the top 28 genes that display transcriptional memory (Figure 1C) revealed that primed genes with the strongest reactivation phenotype in HeLa cells belong to two main gene families: GBPs and HLAs. These gene families form two separate gene clusters on chromosomes 1p22 and 6p21, respectively. The majority of IFN\(\gamma\)-induced genes in the HLA cluster and 3 of 6 IFN\(\gamma\)-induced genes in the GBP cluster show transcriptional memory (Figures S4A and S4B). Moreover, genes of the GBP cluster fall within a single topologically associating domain (TAD) (Figure S5A; Rao et al., 2014). However, not all IFN\(\gamma\)-primed genes appear in clusters. CD74 on chromosome 5q33 is an example of an isolated gene showing IFN\(\gamma\)-mediated transcriptional memory (Figure S4C). Local chromatin domains often show regulatory mechanisms spanning several genes; for instance, at the classic \(\beta\)-globin gene cluster, where a proximal locus control region governs temporally and developmentally regulated expression of globin genes (Noordermeer and de Laat, 2008). Furthermore, another report showed that long non-coding RNAs control expression of genes in a chemokine cluster (Fanucchi et al., 2019). Therefore, despite the lack of long-term maintenance of changes in local chromatin structure (Figure 5), it is possible that longer-range chromatin organization has a selective effect on these clusters and regulates priming and memory.

**Cohesin Inhibits Establishment but Not Maintenance of IFN\(\gamma\) Memory for Most Genes in the GBP and HLA Clusters**

To explore the role of long-range chromatin interactions, we manipulated the levels of cohesin, the principal organizer of local chromatin interactions (Hadjur et al., 2009; Kagey et al., 2010; Rao et al., 2017). We removed functional cohesin specifically during different stages of IFN\(\gamma\) priming and memory, using a HeLa cell line in which both alleles of the essential Kleisin subunit SCC1/RAD21 of the cohesin ring are tagged with an auxin-inducible degron (AID) tag (Wutz et al., 2017; Figure S5B). Combined with the Oryza sativa-derived TIR1 E3 ligase, this degron results in rapid
A

osTIR1+ RNA-seq

protein recovered

SCC1 SCC1

no auxin control

48h window

IFNγ (+-)

priming reinduction

IAA

B

log2 fold change between reinduction and priming

P-adj<0.001

more memory

less memory

GBP genes

HLA genes

top non-cluster memory genes

GBP1 GBP5 GBP4 HLA-DMA CD74 HLA-DMB HLA-DMA CD74

AKNA AKNA

C

420 kbp

CD74 GBP2 GBP1 GBP5 GBP4 GBP5 GBP1

mRNA levels/ 10^3 (AU)

mRNA levels/ 10^3 (AU)

n=3

n=3

D

379 kbp 18.5 kbp 125 kbp

c6orf10/ HCG23/ BTNL2

CD74 GBP2 GBP1 GBP4 GBP5 GBP5 GBP1

mRNA levels/ 10^3 (AU)

mRNA levels/ 10^2 (AU)

n=3

n=3

E

CD74

AKNA

mRNA levels/ 10^3 (AU)

mRNA levels/ 10^2 (AU)

n=3

n=3

(legend on next page)
protein depletion in the presence of auxin (indole-3-acetic acid [IAA]; Nishimura et al., 2009). Based on SCC1-EGFP-AID fluorescence, we determined that auxin mediated depletion of SCC1 is complete in 3 h and fully recovers within 48 h upon IAA washout (Figure S5C).

To determine the role of cohesin in establishment of transcriptional memory, we depleted SCC1 in osTIR1-positive cells during IFNγ priming (Figure 6A) and collected samples for RNA-seq analogous to the transcriptional memory assay as in Figure 1B. Previous work reported that cohesin depletion has a minor effect on steady-state gene expression (Rao et al., 2017; Schwarzer et al., 2017; Wendt et al., 2008; Zuin et al., 2014). However, we found that, during IFNγ gene activation, cohesin loss has broader effects on transcription. 1,628 genes were affected positively and negatively (above 0.5 or below −0.5 log2 fold change cutoff, P-adj < 0.05) by acute short-term SCC1 depletion during IFNγ priming (Figure S5D). These effects are consistent with a recent report of cohesin depletion during LPS activation that showed many of the inducible genes to be affected (Cuatreo et al., 2018). Despite these expected global effects, we found cohesin loss to uniquely affect IFNγ-primed gene clusters where the majority of the clustered memory genes (GBPs and HLAs) showed enhanced memory; i.e., stronger gene expression upon reinduction following IFNγ priming in the absence of SCC1 (Figure 6B). Although cohesin had no bearing on induction, reinduction was enhanced even though cohesin levels were restored to normal levels before the second exposure to IFNγ (Figures 6A, 6C, 6D, and S5C). In contrast, the majority of non-cluster-associated memory genes lost memory upon cohesin depletion or were unaffected (e.g., CD74 and AKNA) (Figures 6B and 6E). The effect on transcriptional memory is specific to SCC1 depletion because IAA treatment of SCC1-EGFP-AID-tagged cells lacking the osTIR1 E3 ligase had no effect on any of the analyzed genes during any stage of IFNγ priming, memory, or reinduction (Figures S6A–S6D). Although all strongly expressed genes in the GBP and HLA clusters showed enhanced memory upon cohesin depletion, a minor subset of genes in the clusters that are expressed at much lower levels did not display such an effect of cohesin depletion (Figures S7A–S7D), indicating that cohesin control is most prominent for robustly expressed genes.

Among the top 15 primed genes we found 3 additional non-clustered genes: GTPBP2, INHBE, and HIST1H2AC (Figure S7E). We confirmed that, among these genes, those with adjacent CD74 (Figure S4C) and AKNA, IFNγ-induced memory is restricted to the identified candidates with no effect on neighboring genes, with a few exceptions of more distal genes; i.e., TMEM268 (proximal to AKNA) and MARS (proximal to INHBE) (Figure S7F). All of these non-cluster associated genes showed impaired memory establishment upon cohesin depletion, contrary to the cluster-associated GBP and HLA genes (Figures 6E, S6D, S7G, and S7H).

Next, to determine whether impaired GBP/HLA gene memory is specifically linked to cohesin loss during IFNγ priming, we primed cells in the presence of cohesin but removed cohesin during maintenance of the primed state (Figure S6E). Despite systemic loss of SCC1, removal of cohesin function during the memory window (as opposed to during priming) had no effect on maintenance of GBP5 or HLA-DRA transcriptional memory (Figure S6F). These results indicate that SCC1 function is selectively required during the priming phase of long-term memory establishment but becomes dispensable when memory is established.

Local, Intra-TAD Cohesin Binding Restricts Transcriptional Memory of the GBP5 and GBP1 Genes

The selective effect of cohesin depletion during IFNγ priming on the maintenance of the primed state led us to the hypothesis that cohesin controls local chromatin structure within the cluster, restricting long-term memory. However, loss of cohesin also has broad effects on gene expression (Figure S5D). It is therefore possible that the effect of cohesin depletion on clustered GBP and HLA genes is indirect. To distinguish between these possibilities, we mapped cohesin binding and locally manipulated its function. We focused on the GBP cluster as the principal locus of IFNγ-induced transcriptional memory. First we performed a ChiP-seq experiment for SCC1 during IFNγ stimulation and washout as outlined in Figure 5A. We identified several cohesin-bound sites, including at the boundaries of the previously identified TAD (Rao et al., 2014; Figures 7A and S5A). Among these, we found three prominent peaks within or immediately adjacent to the GBP genes that show IFNγ priming, which we designated sites: A, B, and C (Figures 7A and 7B). Although global cohesin positions were preserved throughout the experiment, and occupancy did not change considerably, particularly during memory (Figure S7I), we found that two of the sites in the GBP cluster were remodelled upon IFNγ stimulation. Site A on the TAD boundary showed loss of cohesin occupancy during priming, whereas site C within the cluster displayed enhanced chromatin accessibility (as measured by ATAC-seq) at this stage (Figures 7A and 7B). These findings indicate that cohesin is chromatin bound in the GBP cluster and that IFNγ priming results in local changes in cohesin binding or organization.

Next we determined whether local cohesin binding is functionally required for GBP gene priming. We performed CRISPR-Cas9

Figure 6. Cohesin Negatively Regulates Memory for Most Genes in the GBP and HLA Clusters but Not for Genes Outside of Those Clusters

(A) Outline of a transcriptional memory experiment (analogous to Figure 1B) combined with auxin-mediated transient depletion of SCC1 during priming, analyzed by RNA-seq.

(B) HeLa Kyoto SCC1-EGFP-AID osTIR1-positive cells were subjected to the IFNγ and auxin treatment regimen outlined in (A) and processed for RNA-seq. The Log2 fold change for memory genes between reinduction (with or without auxin) and priming (without auxin) was plotted. “0” indicates no memory. The genes shown were selected based on a p-adj value below 0.001 as determined by the DESeq2 software (Love et al., 2014).

(C) Top: representation of the genomic structure of the GBP locus. Bottom: individual gene plots from the data described in (B). Error bars, SD; n = 3 biological replicates.

(D) Top: representation of the genomic structure of the HLA locus. Bottom: data presented as in (C) but for the HLA cluster genes.

(E) Data presented as in (C) but for the CD74 and AKNA genes.

See also Figures S4–S7.
A cohesin sites

TAD boundaries
naive priming
2 days washout
7 days washout

Chromatin accessibility

n p 2d 7d

CRISPR-Cas9 knock-out of cohesin sites
gene expression measurement

B cohesin sites

TAD boundaries
naive priming
2 days washout
7 days washout

Chromatin accessibility

n p 2d 7d

C

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(n≥3)
Mutagenesis to selectively remove the three principal cohesin binding sites in the cluster. We created polyclonal CRISPR-targeted cell populations, deleting an ~1-kb region encompassing each site, and determined the effect of the perturbation on transcriptional memory of the cluster-based GBP1 and GBP5 genes as well as HLA-DRA and a non-cluster memory gene, CD74. We found that removal of the TAD border (site A) or the adjacent intra-TAD (site B) cohesin sites has no significant effect on reinduction of any of the analyzed genes. However, removal of site C, which is also positioned within the TAD, resulted in enhanced priming and stronger reinduction of GBP1 and GBP5. Site C coincides with a known CCCTC-binding factor (CTCF) binding site in HeLa cells (ENCODE: ENCSR000AOA). Importantly, we found this effect to be specific for GBP genes in the cluster in which the cohesin binding site was mutated. HLA-DRA, a gene in a different cluster, and CD74, a non-clustered memory gene, did not show enhanced priming in this mutant. To confirm this result, we isolated a monoclonal CRISPR line for site C and obtained similar results; loss of site C led to enhanced priming and reinduction specifically of the local GBP1 and GBP5 genes (Figure 7C). These results show that, similar to systemic degronomediated cohesin removal, deletion of a local cohesin binding site leads to an enhanced primed state specifically of GBP genes. Recent work reported that decreased enhancer-promoter proximity is required for robust expression of the sonic hedgehog gene during neuronal differentiation (Benabdallah et al., 2019). We speculate that cohesin acts at the GBP and, possibly, HLA clusters in an analogous manner, restricting local contacts and impeding priming and memory establishment.

In summary, we discovered that IFNγ-mediated priming of cells results in a memory of this event that is maintained through numerous cell division cycles, indicating an active self-propagating process. A key insight from our analysis is that strongly primed genes reside in genomic clusters. Memory is controlled, at least in part, by local cohesin function that restricts IFNγ-induced gene expression output. These findings pave the way to explore further mechanisms and ultimately define the link between interferon priming and memory of innate immune signaling.

**STAR METHODS**

Detailed methods are provided in the online version of this paper and include the following:

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### QUANTIFICATION AND STATISTICAL ANALYSIS

**SUPPLEMENTAL INFORMATION**

Supplemental Information can be found online at https://doi.org/10.1016/j.molcel.2020.10.005.

**ACKNOWLEDGMENTS**

We thank Jan-Michael Peters (IMP, Vienna) for sharing HeLa SCC1-AID cells; the flow cytometry facilities of the IGC and Sir William Dunn School of Pathology (University of Oxford) for services and assistance; the EMBL Genomics Core Facility (Heidelberg), Oxford Genomics Centre, and Wellcome Centre for Human Genetics for library preparation and RNA-sequencing; and Moises Mallo, the late Alekos Athanasiadis (both at IGC), and the members of the Jan-Sen Lab for helpful comments and suggestions. This work was supported by Fundação para a Ciência e a Tecnologia (FCT) postdoctoral fellowship SFRH/BDP/117179/2016 (to W.S.) and doctoral fellowship PD/BD/128438/2017 (to S.S.H.T.) and an “Investigador FCT” position (to L.E.T.J.). Research funding was provided by ERC consolidator grant ERC-2013-CoG-615638 and a senior Wellcome research fellowship (both to L.E.T.J.).

**AUTHOR CONTRIBUTIONS**

W.S. and L.E.T.J. conceived the study. W.S. designed and performed the experiments, analyzed the data, and created the figures. J.F.M. provided tissue culture and experimental expertise and assisted with construction of the EGFP::GBP5 cell line. S.S.H.T. generated cohesin site mutants and helped to design the experiments and interpret the data. L.E.T.J. helped to design the experiments and interpret the data and acquired funding. W.S. and L.E.T.J. wrote the manuscript.

**DECLARATION OF INTERESTS**

The authors declare no competing interests.
**SUPPORTING CITATIONS**

The following references appear in the Supplemental Information: Wang et al., 2018.

**REFERENCES**


Molecular Cell

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# STAR Methods

## Key Resources Table

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**RESOURCE AVAILABILITY**

**Lead Contact**
Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Lars E.T. Jansen (lars.jansen@bioch.ox.ac.uk).
Materials Availability
All unique/stable reagents generated in this study are available from the Lead Contact with a completed Materials Transfer Agreement.

Data and Code Availability
The data reported in this paper was deposited in the Gene Expression Omnibus (GEO) database (accession number: GSE150199).
Original data for figures in the paper is available at Mendeley Data http://dx.doi.org/10.17632/86yruz7sf8.2

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Human Cell Lines
- HeLa (female, RRID: CVCL_0030)
- HeLa EGFP::GBP5
- HeLa H3.3-HA (Bodor et al., 2013)
- Hs27 (male, RRID: CVCL_0335)
- HeLa Kyoto (female, RRID: CVCL_1922)
- HeLa Kyoto SCC1-EGFP-AID osTIR1+ (Wutz et al., 2017)
- HeLa Kyoto SCC1-EGFP-AID osTIR1- (Wutz et al., 2017)
- HeLa Kyoto site A cohesin mutant polyclonal population
- HeLa Kyoto site A cohesin mutant polyclonal population 2
- HeLa Kyoto site B cohesin mutant polyclonal population
- HeLa Kyoto site C cohesin mutant polyclonal population
- HeLa Kyoto site C cohesin mutant clonal population
- HCT116 (male, RRID: CVCL_0291)
- hTERT RPE-1 (female, RRID: CVCL_4388)
- HEK-239 (female, RRID: CVCL_0045)

Culture Conditions
All cell lines used were grown at 37°C, 5% CO2. HeLa, male primary fibroblasts (Hs27) and HEK293 cells were grown in DMEM containing glucose, glutamate and pyruvate (GIBCO, 41966-029) supplemented with 10% newborn calf serum (GIBCO, 16010-159) and 1% Penicillin-Streptomycin (GIBCO, 15140-122). HCT116 cells were grown in McCoy’s 5a Medium (Modified) containing glutamine (GIBCO, 26600-023) supplemented with 10% Fetal Bovine Serum (GIBCO, 10500-064) and 1% Penicillin-Streptomycin (GIBCO, 15140-122). hTERT RPE-1 cells were grown in DMEM:F12 Medium (GIBCO, 21331-020) supplemented with 10% Fetal Bovine Serum (GIBCO, 10500-064) and 1% Penicillin-Streptomycin (GIBCO, 15140-122). Cells were counted with Scepter Handheld Automated Cell Counter with 60 μm sensors (Merck, PHCC60050). Transfection of cells was performed using Lipofectamine LTX (Thermo Fisher Scientific) according to the manufacturer’s instructions.

METHOD DETAILS

Reagents
Unless otherwise noted, chemicals used were obtained from Merck and enzymes from New England Biolabs. IFNγ, IFNα and IFNβ were used at a concentration of 50ng/mL. Triptolide was used at concentration of 2nM unless stated otherwise. Auxin: indole-3-acetic acid sodium salt (IAA) was used at concentration of 500μM. The following antibodies were used in this work: GBP5 (Cell Signaling Technology, 67798), Abcam, ab96119, IRF1 (Cell Signaling Technology, 8478), EGFP (Chromotek, 3H9), α-TUB (Merck, T9026), anti-rabbit (fluorophore conjugated) (LI-COR, 926-32211), anti-mouse (fluorophore conjugated) (Rockland, 610-744-124), RNA Pol II (Abcam, ab8119), HA (Merck, 12CAS), H3K27ac (Abcam, ab4729), H3K36me3 (Abcam, ab9050), H3K79me2 (Abcam, ab3594), H3K4me2 (Abcam, ab32356), SCC1 (RAD21) (Abcam, ab992).

FACS and Cytometry
For fluorescence-activated cell sorting (FACS) and cytometry, cells were collected by centrifugation for 5 minutes at 500 g, re-suspended in ice-cold Sorting Medium (1% Fetal Bovine Serum in PBS, 0.25mg/mL Fungizone (Thermo Fisher Scientific), 0.25μg/mL/10μg/mL Amphoterocin B/Gentamicin (GIBCO)) and filtered using 5mL polystyrene round-bottom tubes with cell-strainer caps (Falcon) before sorting (FACSAria) or cytometry analysis (FACSCalibur) (BD Biosciences). For sorting, cells were collected in Conditional Medium (1:1 mixture of fresh complete medium and medium collected from proliferating cell cultures that is 0.45μm filtered, supplemented with 20% Fetal Bovine Serum, 0.25mg/mL Fungizone (Thermo Fisher Scientific), 0.25μg/mL/10μg/mL Amphoterocin B/Gentamicin (GIBCO)).
DNA Constructs and Genome Engineering

To construct the EGFP::GBP5 promoter trap cell line the plasmid pX330-U6-Chimeric_BB-CBh-hSpCas9 (Addgene #42230) (Cong et al., 2013) was employed with two guide RNA (gRNA) sequences: 5′-GCAAGATACATCTAGACA-3′ and 5′-GGCA- CATGGGTCTGACATG-3′ targeting exon 2 of the GBP5 gene that encodes the start codon. The gRNAs were selected using the GPP sgRNA Designer software (Doench et al., 2016). The homology repair template consisted of an EGFP cassette generated from a synthetized DNA EGFP-FLAG-miniAID-FLAG (GeneScript) with 75bp homology arms to the GBP5 gene added during PCR amplification (Q5 polymerase, NEB). The homology arms span: chr1:89269477–89269552 and chr1:89269555–89269630 of the human hg38 genome. For both guide RNAs the homology arms were designed to introduce a silent mutation in the protospacer-adjacent motif (PAM) recognition sequence after successful repair to prevent Cas9 re-cutting. After transfection of the mixture of the plasmid containing the Cas9/guide RNA and the homology repair template (1:2 ratio) the cell lines treated with different guide RNAs were mixed and grown for 48h. Cells were induced with INFγ and a polyclonal population was sorted based on EGFP fluorescence as described above. After 48h of proliferation without INFγ a polyclonal population was again sorted based on the lack of EGFP signal. This step was performed to remove cells with an improper targeted EGFP cassette. The cells were left to proliferate for 48h, induced again with INFγ and single cell sorted to generate monoclonal lines. Following clonal selection, cells were maintained in culture for about 30 days to erase INFγ priming before being used in experiments.

To construct the GBP gene cluster cohesin site mutants we deleted a ~1 kb region using a double Cas9 cut strategy. To achieve this we used a combination of two plasmids: lentiCRISPR v2 (Addgene #52961) (puromycin resistance) and lentiCRISPR v2-Blast (Addgene #83480) (blasticidin resistance) (Sanjana et al., 2014) with gRNAs flanking the selected cohesin sites. The double drug resistance allowed us to select for a dual Cas9 cut on either site of the cohesin site. Two independent guide RNA pairs for the A site (hg38, chr1:88,997,800–88,999,895) were: 5′-AAATAGTATATCAAAAGAGG-3′, 5′-GTCTCAATTGCTGTGCTGTG-3′ and 5′-TAGATCCCTAGAAGCTTTCA-3′. Guide RNAs for the A site (hg38, chr1:88,997,800–88,999,895) were: 5′-AAATGCTTATGAGTGG-3′ and 5′-GTCTCCTCAATAGACCTTG-3′. Guide RNAs for the C site (hg38, chr1:89357816–89359284) were: 5′-ATTCCATCTCGTCTAGGCG-3′ and 5′-ACATGCTTATGCGATACCC-3′. The gRNAs were selected using the GPP sgRNA Designer software (Doench et al., 2016). The plasmids with cloned gRNAs were co-transfected (as described above) with lentiviral packaging plasmids: psPAX2 (Addgene #12260), pMD2.G (Addgene #12259) into HEK293 cells at a molar ratio of: 4:3:1. 24h later, medium was refreshed and the cells were incubated for 3 days at 37°C. Medium was collected and filtered through a 0.45 μm filter to obtain the viral particles. For lentiviral infections HeLa cells were incubated in medium with 8 μg/mL of polybrene (Merck) for 1h and infected with a 1:1 mixture of viruses carrying Cas9 with gRNAs flanking a given cohesin site. The cells were left to grow for 48h followed by selection with both blasticidin (1 μg/mL) and puromycin (1 μg/mL). For the site C mutant a monoclonal line was additionally established by FACS.

Transcriptional Memory Assay

Cells were cultured and split into two parallel cultures. One was mock treated with medium and the other with INFγ for 24h unless stated otherwise. Cells were then washed three times with PBS and trypsinized (GIBCO) to remove residual INFγ and cleave the extracellular domains of plasma membrane proteins, including the INFγ receptor, to stop residual signaling. Fresh medium was added and cells were allowed to proliferate for 48h hours unless stated otherwise. Next INFγ was added to the primed and naive cells. After 24h cells were washed, trypsinized, centrifuged 500 g for 5 minutes and the pellets were processed for downstream analysis.

RT-qPCR

Cell pellets (8*10⁶ cells) were re-suspend in 0.25mL of PBS and 1mL of TRIzol Reagent was added per sample. Cells were lysed by extensive pipetting and incubated for 5 minutes at room temperature. Next, 0.2mL of chloroform was added per sample, mixed and incubated for 3 minutes followed by centrifugation at 12000 g for 15 minutes at 4°C. The aqueous phase was mixed with 0.5mL of 100% isopropanol and incubated at room temperature for 10 minutes followed by centrifugation at 12000 g for 10 minutes at 4°C. The supernatant was removed and the pellet was washed with 1mL of 75% ethanol and air-dried for 10 minutes. Next the RNA pellet was re-suspended in 100 μL of RNase-free water and incubated at 60°C for 10 minutes. The residual DNA was removed with DNase I (Roche) and the RNA was purified with RNeasy Mini kit (QIAGEN) according to manufacturer’s instructions. cDNA was prepared using a High-Capacity RNA-to-cDNA Kit (Applied Biosystems) and the libraries were diluted 10 times before qPCR measurements. The qPCR assay was performed with PerfeCta SYBR Green FastMix ROX (Quanta) using primers (at 300nM concentration): ACTB_F: 5′-CTCTTCCAGCCTCCTCTCCT-3′, ACTB_R: 5′-AGCAGCTGTTGGGCTACG-3′, GBP5_F: 5′-TCATATTGCGGCCGTCTG-3′, GBP5_R: 5′-AGGCACTGTTCAAGTGGG-3′, HLA_DRA_F: 5′-GAAAGGACTCTTCAATCAGC-3′, HLA_DRA_R: 5′-AGAGGACGTTGGATGAT-3′, GBP1_F: 5′-GTGGAACTGTTGAAAGCTA-3′, GBP1_R: 5′-CAACTGGACCATTTGCTCTTC-3′, CD74_F: 5′-TGGGAGGTCTGACATG-3′, CD74_R: 5′-ACATTGCTATTGGCATACCTC-3′. All experiments were performed in technical and biological triplicates, for every primer pair a calibration curve was determined and a melting curve was measured at the end of the reaction. The qPCR conditions were as follows: 95°C 3 minutes; [95°C 10 s; 59°C 30 s]x50 cycles. From the calibration curve a linear regression was established and the parameters were used to determine the nucleic acids amount according to the equation: N = 10^(-((Ct - b)/a)), N = relative measure of DNA amount; a = slope of regression; b = y intersect point; Ct = qPCR measurement value.

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**Immunoblotting**

Cell pellets (8×10^6 cells) were re-suspended in protein loading buffer (125mM Tris-HCl pH 6.8, 10% Glycerol, 1% SDS, 0.2% (w/v) Orange G, 5% β-mercaptoethanol) and incubated at 95°C for 5 minutes. Benzonase (50U) was added to the lysates and the samples were incubated in room temperature for 30 minutes. Next, sample concentration was normalized based on the 260nm absorbance determined by NanoDrop. Normalized samples were separated on a 12% SDS-PAGE gel (Bio-Rad), transferred to nitrocellulose (Bio-Rad), blocked with Odyssey Blocking Buffer (LI-COR) and incubated overnight with primary antibodies in Odyssey Blocking Buffer. Blots were then washed three times with TBST (20mM Tris-HCl pH 7.5, 150mM NaCl, 0.1% Tween 20) and incubated with secondary antibodies (LI-COR) in Odyssey Blocking Buffer. Blots were next washed three times with TBST and analyzed by Odyssey Imaging System (LI-COR).

**RNA-seq**

For transcriptional memory experiments in HeLa cells, three biological replicates of naive, priming, primed and reinduced cells were processed for RNA isolation as described above for RT-qPCR measurements. Qubit RNA BR Assay Kit (Thermo Fisher Scientific) was used for quantification and RNA ScreenTape (Agilent) for accessing RNA integrity. RNA was then used for polyA positive library preparation with the NEBNext Ultra RNA Library Prep Kit for Illumina (New England Biolabs), according to the manufacturer’s protocol. Sequencing was performed on the NextSeq 500 System with the NextSeq 500/S50 v2.0 Kit (Illumina) with 75bp read length in single end mode. Library preparation and sequencing was performed at the EMBL Genomics Core Facility.

For the cohesin depletion experiments, HeLa Kyoto SCC1-AID osTIR1 positive / negative cells (Wutz et al., 2017) were treated or not with axuin for 3h followed by treatment with IFNγ (Merck) or mock medium for 16h. Cells were then washed three times with PBS and trypsinized (GIBCO) to remove residual INFγ and cleave the extracellular domains of plasma membrane proteins, including the INFγ receptor, to stop residual signaling. Part of the culture treated with IFNγ was processed for RNA isolation. Fresh medium was added to the other cultures and cells were allowed to proliferate for 48h. Next IFNγ was added or not to the primed and naïve cells. After 24h cells were washed, trypsinized, centrifuged 500 g for 5 minutes and the pellets were processed for downstream analysis. Three biological replicates of naive, priming, primed and induced cells were processed for RNA isolation using the TRIzol Reagent. Qubit RNA BR Assay Kit (Thermo Fisher Scientific) was used for quantification and RNA ScreenTape (Agilent) for accessing RNA integrity. Purification of mRNA, generation of double stranded cDNA and library construction were performed using NEBNext Poly(A) mRNA Magnetic Isolation Module and NEBNext Ultra II Directional RNA Library Prep Kit (New England Biolabs) with previously reported barcode tags (Lamble et al., 2013). The concentrations used to generate the multiplex pool were determined by Quanti-IT PicoGreen dsDNA Assay Kit (Thermo Fisher Scientific). The material was sequenced on the NovaSeq6000 instrument (Illumina) with 150bp read length in paired end mode. Library preparation and sequencing was performed in the Oxford Genomics Centre, Wellcome Centre for Human Genetics.

For both the experiments mapping was performed with bwa-mem software (Li, 2013) version 0.7.17.1 in Simple Illumina mode to the hg38 human genome assembly. Gene readcounts were estimated with htseq-count software (Anders et al., 2013) version 0.9.1. Parameters: union mode, feature type exon, ID Attribute gene_id. DESeq2 version 2.11.40.6 was used to estimate the differential expression between samples (Love et al., 2014). Note that RNA-seq data is presented as log2 fold change to estimate the expression difference. The log2 fold change values represent the multiplicative factor “x” on a log2 scale, convertible to a linear fold change by 2^x. e.g., a log2 fold change of 0.5 equals 2^0.5 = 1.4.

**Single Cell RNA-seq**

For the single cell RNA-seq experiment, naive, stimulated or re-stimulated HeLa cells were processed for single cell sorting (one 96 well plate per condition) as described above, with an additional viability staining using SYTOX AADvanced (Thermo Fisher Scientific). Viable single cells were collected to Lysis Buffer (0.8% Triton X-100, 2U/μl RiboLock (Thermo Fisher Scientific)). The material was used for the single cell Smart-Seq2. Nextera XT library preparation according to the original protocol (Picelli et al., 2013) and sequenced on the HiSeq4000 System (Illumina) with 75bp read length in paired end mode. Library preparation and sequencing was performed in the Oxford Genomics Centre, Wellcome Centre for Human Genetics. Mapping was performed with bwa-mem software version 0.7.17.1 in Simple Illumina mode (Li, 2013) to the hg38 human genome assembly. Gene readcounts were estimated with htseq-count software version 0.9.1 (Anders et al., 2015). Parameters: union mode, feature type exon, ID Attribute gene_id. Gene readcounts were corrected with the following procedure: corrected gene readcounts = gene readcounts / (total mapped reads / 1000000).

**ATAC-seq**

The ATAC-seq procedure was adopted from Chen et al. (2016). In brief, 5×10^6 naive, stimulated or primed cells were collected by centrifugation for 5 minutes at 500 g. Next, the cells were re-suspended in 50μL of ATAC lysis buffer (10mM Tris-HCl pH 7.4; 10mM NaCl; 3mM MgCl2; 0.01% Igepal CA-630) and centrifuged immediately for 10 minutes at 500 g at 4°C. The cell pellets were re-suspended in 50μL tagmentation reaction buffer (25μL 2 × TD buffer, 22.5 μL dH2O, and 2.5μL TDE1 (Tn5 enzyme) (Illumina) and incubated in 37°C for 30 minutes. DNA was purified with MinElute PCR Purification Kit (QIAGEN) (10 μL elution).

For preparation of sequencing libraries the purified DNA was amplified using Q5 Hot start DNA polymerase using indexing primers described in Buenrostro et al. (2013). Initial amplification was performed: 98°C 30 s; [98°C 10 s; 63°C 30 s; 72°C 1 minute]x 5 cycles
followed by library quantification with qPCR: 95 °C 3 minutes; [95 °C 10 s; 58 °C 30 s; 72 °C 1 minute] and additional PCR amplification (conditions as for the initial 5 cycles). The required number of additional cycles was calculated from the qPCR data by determining the number of cycles to reach 50% of the maximum signal. After library amplification and indexing, DNA was purified and size selected with AMPure XP magnetic beads (Beckman Coulter) according to manufacturer’s protocol. After purification the libraries concentration was determined by Qubit dsDNA HS Assay Kit (Thermo Fisher Scientific) according to manufacturer’s protocol. The fragment size was estimated by DNA ScreenTape (Agilent) and the final library quality was measured with KAPA Library quantification kit (Roche) according to manufacturer’s protocol. For sequencing, multiplexed libraries were diluted to 2nM concentration (calculated based on the KAPA Library quantification kit). The sequencing was performed on a NextSeq 550 System with the NextSeq 500/550 v2.5 Kit (Illumina) with 75bp read length in single end mode. The data was mapped to the human genome version hg38 using bowtie2 software (Langmead and Salzberg, 2012) version 2.3.4.3 in default mode. Coverage bigwig files were generated using bamCompare software (Ramírez et al., 2016) against the input sequencing by subtraction, with 50bp bin size and read count scaling.

**ChIPmentation**
The ChIPmentation procedure was adopted from Schmidl et al. (2015). In brief, $12 \times 10^7$ naive, stimulated or primed cells were collected by centrifugation for 5 minutes at 500 g. Next, the cells were re-suspended in 25 mL of medium supplemented with 1% formaldehyde and incubated for 10 minutes in room temperature. Cross-linking was quenched by addition of 25mL of 125mM glycine in PBS. Cells were harvested by centrifugation, washed two times with 10mL of cold PBS, frozen in liquid nitrogen and kept in –80 °C. To fragment chromatin, cell pellets were re-suspended in Lysis Buffer (50mM Tris-HCl pH 7.5; 1% SDS; 10mM EDTA; protease inhibitors (Roche)) and sonicated with Qsonica Q800R2; $10^7$ cells in 0.3mL of buffer per 0.5mL tube with the following conditions: time – 5 minutes, pulse – 30 s ON/OFF, amplitude – 75%. The resulting material was tested for proper sonication by fragment analysis confirming DNA size between 500 and 4000bp. Chromatin samples were aliquoted at $5 \times 10^6$ cells per tube and frozen in –80 °C. For a single immunoprecipitation, 25 μL of protein A or G Dynabeads beads (Thermo Fisher Scientific) were washed twice with ChIP Dilution Buffer (20mM Tris-HCl pH 7.5; 0.1% SDS; 1% Triton X-100; 150mM NaCl; 2mM EDTA; protease inhibitors (Roche)). In parallel, the sample was diluted with ChIP Dilution Buffer up to 2mL and clarified at 12000 g for 10 minutes in 4 °C. Input DNA was collected from the supernatant (10μL). Samples were incubated with the beads and a given antibody over night at 4 °C with rotation. Beads were washed at 4 °C as follows: twice with Low Salt Wash Buffer (50mM Tris-HCl pH 7.5; 150mM NaCl; 1mM EDTA; 1% Triton X-100; 0.1% Na-Deoxycholate; protease inhibitors (Roche)); twice with High Salt Wash Buffer (50mM Tris-HCl pH 7.5; 500mM NaCl; 1mM EDTA; protease inhibitors (Roche)); twice with LiCl Wash Buffer (10mM Tris-HCl pH 7.5; 0.5% NP-40; 0.5% Na-Deoxycholate; 250mM LiCl; 1mM EDTA; protease inhibitors (Roche)); twice with TE Buffer (10mM Tris-HCl pH 7.5; 1mM EDTA; protease inhibitors (Roche)) and finally twice with 10mM Tris-HCl pH 7.5. Beads were re-suspended in 30 μL of ChIPmentation reaction buffer (10 mM Tris-HCl pH 8.0; 5mM MgCl2; 1 μL TDE1 (TnS enzyme) (Illumina). In parallel 1 μL of input was diluted in 29 μL of ChIPmentation reaction buffer. The samples were incubated in 37 °C for 10 minutes followed by washing twice in ice-cold LiCl Buffer and TE Buffer. Next, the beads were suspended in 200μL of Elution Buffer (50mM Tris-HCl pH 7.5; 1% SDS; 10mM EDTA; protease inhibitors (Roche)). In parallel the input reaction was mixed with 170 μL of the Elution Buffer. All samples were incubated at 65 °C overnight for decrosslinking followed by treatment with PureLink RNaseA (Thermo Fisher Scientific) and Proteinase K. The DNA was purified with MiniElute PCR Purification Kit (QIAGEN) (10 μL elution). Library preparation and sequencing was performed as described above for ATAC-seq.

**QUANTIFICATION AND STATISTICAL ANALYSIS**

RT-qPCR and RNA-seq data were collected in triplicate. Standard deviation is reported.

Statistical analyses and P value calculations for the RT-qPCR data were performed by two-tailed unpaired t tests (GraphPad Prism 6.01) and can be found in the figure legends.

RNA-seq data statistics were calculated by the DESeq2 software (Love et al., 2014); pvalues are calculated by the Wald test, padj values are calculated by the Benjamini-Hochberg method.
Supplemental Information

Activation of Clustered $\text{IFN}_\gamma$ Target Genes Drives Cohesin-Controlled Transcriptional Memory

Wojciech Siwek, Sahar S.H. Tehrani, João F. Mata, and Lars E.T. Jansen
Supplementary Figure 1. Transcriptional memory is cell type dependent and is not a consequence of cell selection. Related to Figure 1. (A) HeLa cells were primed and reinduced according to the regime outlined in Figure 1B. HLA-DRA mRNA levels were quantified by RT-qPCR and normalized to ACTB expression. Error bars SD, n=3 biological replicates. (B) Average read counts for three replicate RNA-seq experiments as outlined in Figure 1B were
assembled for each gene for naïve cells and after priming. The log2 fold change of gene read count following priming over those from naïve cells is plotted. “0” indicates no change while positive or negative values indicate responsiveness to IFNγ. Data was ranked according to the mean expression level for both of the conditions and all replicates. Red dots represent genes with an adjusted P value below 0.1. (C) HCT116 and hTERT RPE-1 cells were subjected to IFNγ treatment regime as outlined in Figure 1B, processed for western blotting and probed for GBP5 protein expression. α-tubulin (α-TUB) loading control. (D) Experimental outline to assay the effect of IFNγ exposure on HeLa cell proliferation. (E) Results from experiment shown in panel D. Cells were primed with IFNγ for 24h and left to proliferate for 48h analogous to experiments in Figure 1B, followed by counting of live cells. The results were normalized to naïve cells. Error bars SD, n=3 biological replicates. (F, G) Single cell RNA-seq data from the experiment shown in Figure 3B for HLA-DRA (F) and ACTB (G) in naïve (N=90), priming (N=89) and reinduction (N=91) states. Every dot represents one cell. (H) Experiment to determine cytokine-specific induction of the EGFP::GBP5 promoter trap knock-in cells. Cytometry measurements of EGFP expression before and after stimulation with IFNγ, α and β. Cell frequencies as a function of EGFP fluorescence intensity are plotted.
Supplementary Figure 2. HeLa cell population is uniformly responsive to IFNγ and transcriptional output correlates with establishment of transcriptional memory for the GBP5 gene. Related to Figure 3. (A) Representation of the single cell RNA-seq data in HeLa
cells as in Figure 3C for naïve and IFNγ exposed cells for indicated IFNγ inducible genes. Every dot represents the expression level for the indicated gene for one cell in naïve (N=90) and priming states (N=89). (B) HeLa EGFP::GBP5 cells were subjected to IFNγ treatment regime as outlined in the top panel. After priming, a sample from 20% of top expressing, 20% of bottom expressing and all cells were isolated by fluorescence-activated cell sorting (FACS) and left to proliferate for 48h. Cells were then reinduced or not with IFNγ and subjected to cytometry to assay EGFP expression. Cell frequencies as a function of EGFP fluorescence intensity are plotted. Cut-off for cell percentages expressing or not expressing are based on the “all cell” sort IFNγ stimulation.
Supplementary Figure 3. Absence of ongoing transcription during maintenance of transcriptional memory. Related to Figure 4. (A) EGFP::GBP5 reporter cells were subjected to IFNγ treatment regime as outlined in Figure 4A and processed for cytometry. Cell frequencies as a function of EGFP fluorescence intensity are plotted. Red dotted lines are fiducial marks to aid comparison. (B) HeLa cells were subjected to IFNγ treatment regime and
RNA-seq as outlined in Figure 1B, C. Expression levels of genes that show elevated mRNA in primed over naïve cells (see Figure 4C) were plotted (Error bars SD, n=3 biological replicates). (C) Outline of experiment to determine the minimal inhibitory concentration of triptolide impacting expression of IFNγ responsive genes. (D) HeLa cells were subjected to IFNγ and triptolide titration series as outlined in panel C, processed for western blotting and probed for an IFNγ early response gene – IRF1 expression. α-tubulin (α-TUB) loading control.
Supplementary Figure 4. The two major classes of genes that are primed for re-expression, GBPs and HLAs, reside in genomic gene clusters. Related to Figure 6. (A) Representation of the genomic structure of the GBP locus and the processed RNA-seq data from experiment shown in Figure 1B. Read counts were assembled for each gene of the GBP
cluster (green) and three flanking genes (black) following priming and reinduction. (B) As in A, but for the HLA gene cluster. (C) As in A, but for the non-cluster CD74 gene. Error bars SD, n=3 biological replicates.
Supplementary Figure 5. Auxin-mediated depletion of the cohesin subunit SCC1 is complete in 3 hours, fully recovers within 48 hours and has broad effects on gene expression. Related to Figure 6. (A) Published Hi-C dataset showing GBP genes clustered in a single topologically associating domain (TAD) in the GM12878 cell line with 5kbp resolution.
(Rao et al., 2014), the data was visualized with the 3D Genome Browser (Wang et al., 2018).

(B) Schematic of SCC1 genomic structure in HeLa Kyoto SCC1-EGFP-AID cell line. Exon 14 bearing the end of the SCC1 ORF is tagged in frame with EGFP and an AID-degron. Both alleles are modified (Wutz et al., 2017).

(C) Cytometry analysis of EGFP expression in the SCC1-EGFP-AID cells after 3h depletion with auxin (IAA) and following 24h and 48h washout of auxin. Cell frequencies as a function of EGFP fluorescence intensity are plotted.

(D) Representation of processed RNA-seq data from HeLa Kyoto, SCC1-AID, osTIR1 positive cells as described in upper left panel (identical to Figure 6A). Average read counts for three replicate experiments were assembled for each gene following control (-IAA) or auxin treatment (+IAA). The log2 fold change of gene read counts after IAA treatment over mock treated cells is plotted following priming, in primed cells and after reinduction. “0” indicates no change while positive or negative values indicate increased or decreased expression upon IAA treatment, respectively. Data was ranked according to the mean expression level for both of the conditions and all replicates. Red dots represent genes with an adjusted P value below 0.1.
Supplementary Figure 6. Effects on establishment of transcriptional memory following auxin addition to HeLa SCC1-AID cells are dependent on cohesin depletion; and cohesin is not required for maintenance of transcriptional memory of the GBP5 and HLA-DRA genes. Related to Figure 6. (A) Outline of a transcriptional memory experiment (analogous to

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Figure 6A) but in cells lacking the osTIR1 E3 ligase, analyzed by RNA-seq. (B) HeLa Kyoto SCC1-EGFP-AID osTIR1 negative cells were subjected to IFNγ and auxin treatment regime as outlined in panel A. mRNA levels of GBP cluster genes were quantified by RNA-seq in auxin or mock treated cells. Error bars SD, n=3 biological replicates. (C) Data presented as in B, but for the HLA cluster genes. (D) Data presented as in B, but for the CD74 and AKNA non-cluster genes. (E) Outline of a transcriptional memory experiment (analogous to Figure 6A) combined with auxin-mediated transient depletion of SCC1-EGFP-AID in HeLa Kyoto osTIR1 positive cells. Cells were treated with auxin to remove SCC1 protein, or left untreated after priming but before reinduction (during the memory window). (F) Cells were subjected to IFNγ and auxin treatment regime as outlined in E and processed for RT-qPCR of GBP5 and HLA-DRA, normalized to ACTB expression. Error bars SD, n=3 biological replicates.
Supplementary Figure 7. Non-cluster genes, GBP genes at cluster borders and some HLA cluster genes lack enhanced memory establishment after cohesin depletion. Related to Figure 6 and 7. (A) HeLa Kyoto, SCC1-AID, osTIR1 positive cells were subjected to IFNγ treatment regime as outlined in Figure 6A. mRNA levels of indicated GBP genes were...
quantified by RNA-seq. Error bars SD, n=3 biological replicates. (B) As in A, but for HeLa Kyoto, SCC1-AID osTIR1 negative cells. (C, D) as in A and B, but for indicated HLA cluster genes. (E) Top memory genes as identified in the HeLa Kyoto, SCC1-AID, TIR1 positive cells by DESeq2 software based on experiment in Figure 6A. Genes not associated in the GBP or HLA clusters are indicated in red. Log2(FC) – log2 fold change between priming and reinduction. P-adj – adjusted P value. (F) Representation of processed RNA-seq data from experiment shown in Figure 6A for top non-GBP or -HLA cluster memory genes. Read counts were assembled for indicated genes (labeled in green) and three flanking genes (labelled in black) following priming and reinduction. The experiment was performed in biological triplicate in HeLa SCC1-EGFP-AID osTIR1 positive cells without IAA addition. Error bars SD. (G) HeLa Kyoto, SCC1-AID, osTIR1 positive cells were subjected to IFNγ and IAA treatment regime as outlined in Figure 6A. Plotted mRNA levels were quantified by RNA-seq (Error bars SD, n=3 biological replicates). (H) As in G but for HeLa Kyoto, SCC1-AID osTIR1 negative cells. (I) Heat map analysis of SCC1 genome-wide occupancy in naïve cells, after priming and at 2 and 7 days after IFNγ washout. SCC1 binding sites were ranked based on SCC1 signal in naïve cells. False color scale represents SCC1 occupancy corrected for input.